

Entomopathogenic Fungus, *Isaria fumosorosea*, and Aphid Parasitoid, *Lysiphlebus testaceipes*, -Managing Infestations of Brown Citrus Aphid, *Toxoptera citricida*

David Pick^{1,2}, Pasco B. Avery², Wayne B. Hunter³, Charles A. Powell², David G. Hall³

¹Florida Atlantic University, Harriet L Wilkes Honors College, 5353 Parkside Drive, Jupiter, FL 33458, USA

²University of Florida, Institute of Food and Agricultural Sciences, Indian River Research and Education Center, 2199 South Rock Road, Fort Pierce, FL 34945, USA

³USDA, ARS, U. S. Horticultural Research Laboratory, Subtropical Insect Research Unit, 2001 South Rock Road, Ft. Pierce, FL 34945, USA



In Florida, the continual use of broad-spectrum chemical insecticides has concerned growers in the citrus industry since “green” produce is viewed as healthier and more desirable to the consumer. Bio-insecticides, such as PFR 97 (*Isaria fumosoroseus* strain Apopka 97 [*Ifr*], [Fig. 1.5]) may offer an alternative to growers, and minimize ecological impact on non-target organisms (Openders et. al 2004). This study assessed *Ifr*'s effects on *Lysiphlebus testaceipes* (Cresson) (Fig. 1.3), a non-target parasitoid of the citrus pest *Toxoptera citricida* (Kirkaldy), (Fig. 1.2).

MATERIALS & METHODS

- Four groups of citrus were labeled 1-4 (five carrizo citrus [25-30 cm tall] per group) for control (water only), *L. testaceipes* only, *Isaria fumosorosea* (*Ifr* only), and *L. testaceipes* plus *Ifr*, respectively, and kept in bugdorm cages (35 cm³) at 24±0.03 °C, 45±0.29 % RH, and 16L:8D hr photoperiod supplied by four 40W Philips full spectrum fluorescent tubes (Figure 1.1).
- Twice, aphids were added to each cage; the 1st set was added the same day as cage set up the 2nd set was added two weeks later (each time four adult alate aphids were added per plant).
- Two times, *L. testaceipes* adults were added to cages 2 & 4 the 1st set was added five days after the final aphid release the 2nd set was added one week later (each time two adult parasitoids were added)
- Prior to spraying *Ifr* *L. testaceipes* adults were removed
- Isaria fumosorosea* was sprayed using a 180 ml sterile Nalgene® aerosol spray bottle (1 g PFR 97 20% WDG Certis [USA]/100 ml autoclaved de-ionized water).
- After the cage was sprayed *L. testaceipes* adults were replaced in their respective cages, and one additional adult was added to each plant.
- Data was recorded for the numbers of aphids, mummies, emerged parasitoids (mummies with holes) and dead aphids in each cage every day (except weekends) for 22 days.
- This experiment was conducted twice. (experiments 1 & 2)



Figure 1.1 From left to right cages 1-4: citrus aphids treatments control (water only), *Lysiphlebus testaceipes* only, *Isaria fumosorosea* (*Ifr* only), and *L. testaceipes* plus *Ifr*, respectively



Figure 1.2 A brown citrus aphid infestation on Carrizo Citrus



Figure 1.3 A *Lysiphlebus testaceipes* adult



Figure 1.4 A brown citrus aphid (mummy) killed by a now adult (emerged) *Lysiphlebus testaceipes* larva



Figure 1.5 *Isaria fumosorosea* infection on an adult apterous aphid

RESULTS

- The fungal treatment did not have an apparent negative influence on parasitism rates even though in experiment one, significantly more aphids were successfully parasitized in the fungus-treated cage than in the non-fungus-treated cage this is considered a positive not a negative influence; however, in experiment 2 these numbers were not significantly different (figs. 2.1 & 2.2).
- No significant difference was observed in the emergence rate between mummies in the presence of fungus and the non fungus treatment (figs 2.3 & 2.4).
- Toxoptera citricida* mortality rates were significantly higher in both parasitoid treatments, often nearly 100%, indicating *L. testaceipes* is a highly effective aphid BioControl agent even in the presence of *Ifr*.

DISCUSSION

- Isaria fumosorosea* and *L. testaceipes* were shown to be compatible biological control agents against *T. citricida*. The use of PFR 97 20% WDG will not likely interrupt the efficient naturally occurring control of *T. citricida* by *L. testaceipes*, and should be able to be used in citrus production without concern towards *L. testaceipes* (a big plus for citrus growers). These findings with *L. testaceipes* open new avenues for future IPM compatibility research.
- Although *Ifr* was a disappointingly ineffective manager of *T. citricida* during these experiments, an ~10 viable blastospores per mm were deposited by the sprayer, and *Ifr* was detected in treatments 3 & 4 up until the last day. This study did not endeavor to evaluate the efficacy of *Ifr* on *T. citricida* since this has been done (Poprawski et. al 1999). One potential explanation for *Ifr*'s limited effectiveness against *T. citricida* in this study is that dead aphids do not stay on their host plant. They fall off taking the *Ifr* inoculum with them, potentially slowing down would be epizootics. In which case, having another highly mobile insect such as *L. testaceipes* inoculating *T. citricida* individuals with *Ifr* might increase *Ifr*'s effectiveness. In fact, a higher mortality rate in treatment 4 (*Ifr* with *L. testaceipes*) was often observed. Under these considerations, the use of PFR 97 20% WDG may offer a greener alternative to citrus growers, and minimize ecological impact on non-target organisms.

REFERENCES

- Openders koul, G. S. Dhaliwal, Gerrit W. Cuperus. 2004. Integrated Pest Management. CABI Publishing. 138.
- Poprawski, T.J., Parker, P.E. & Tsai, J.H. (1999) Laboratory and Field evaluation of hyphomycete insect pathogenic fungi for control of brown citrus aphid (Homoptera:Aphididae). Environmental Entomology 28, 315-321.

Photo Credits:

Background, Figs. 1.3-1.5: David A. Pick; Figs. 1.1-1.2: Christine Lynch

Acknowledgements

Although, I designed much of this project with input from Drs. Avery, Hunter, and Powell, it was Christine Lynch who collected data and made key protocol modifications. Her efforts led to the success of the experiment, and I thank her for the contributions she made.

Thank you Brandon Paradise, Robin Barnes, and Gail Amafitano for your work counting aphids. Thank you Dr. Ronald Cave and Dr. William Overholt for the use of your bug dorms. Thank you Phyllis Rundell, Eliza Duane, Lindsay Brock for your help in brainstorming. Thank you, Deanna Pick without your support and encouragement this study would not have occurred. Thank you Maria Gonzalez for your help transporting Materials. Thank you Anna Sarah Hill for rearing the aphid colonies.

